

## DUB assay explorer panel

UbiQ code : UbiQ-L03

Amount : 4 x 25 µg lyophilized powder

Purity : ≥95% (purified by RP-HPLC)

Storage : upon arrival, powder at –20°C, solution at –80°C. Please avoid multiple freeze/thaw cycles and protect from light.

## Productsheet

**Background.** UbiQ-L03 is a panel of 4 deubiquitylating enzyme (DUB) activity assay reagents (25 ug each) prepared by total chemical synthesis.<sup>1</sup>

- **Ub-AMC (UbiQ-001)** is a quenched fluorogenic DUB substrate based on the C-terminal derivatization of ubiquitin with 7-amido-4-methylcoumarin (AMC). Cleavage of the amide bond between Gly76 of ubiquitin and the AMC moiety results in an increase in fluorescence at  $\lambda_{\text{Ex}}/\lambda_{\text{Em}} = 380/460$  nm.<sup>2</sup>
- **Ub-Rh110Gly (UbiQ-002)** is a quenched fluorogenic DUB substrate based on the C-terminal derivatization of ubiquitin with rhodamine110-Gly (Rh110Gly). Cleavage of the amide bond between Gly76 of ubiquitin and Rh110Gly results in an increase in fluorescence at  $\lambda_{\text{Ex}}/\lambda_{\text{Em}} = 535/485$  nm.<sup>3</sup>
- **TAMRA-Lys(Ub)-Gly (UbiQ-012)** is a fluorescence polarization assay reagent based on a 5-carboxytetramethylrhodamine (TAMRA  $\lambda_{\text{Ex}}/\lambda_{\text{Em}} = 530/580$  nm) modified Lys-Gly sequence that is linked to ubiquitin via a native isopeptide bond with the lysine side-chain.<sup>4</sup>
- **Ub-aminoluciferin (UbiQ-036)** is a DUB substrate based on the C-terminal derivatization of ubiquitin with 6-aminoluciferin (Luc). Upon cleavage by a DUB, the released 6-aminoluciferin functions as a substrate for luciferase, allowing detection of luminescence as a read-out for DUB-activity.<sup>5</sup>

### important - Sample preparation.

- dissolve the powder in as little DMSO as possible (e.g. 25 mg/mL = 25 ug in 1 uL DMSO)
- add this DMSO stock slowly to milliQ (please note the order of addition)
- next buffer as desired (e.g. with 1M HEPES to 50 mM HEPES). In general, HEPES and Tris buffers are standard for DUB assays
- for example, a stock of 100 uM in buffer ( $\pm 0.9$  mg/mL,  $\pm 4$ vol% DMSO) is diluted 1000x to prepare a final standard assay solution of 100 nM substrate concentration
- please note that certain DUBs react different to low or high NaCl concentrations.
- TCEP or DTT can be used as reducing agent for the DUB (see Wrigley et al. *Cell Biochem Biophys* **2011**, 60, 99).

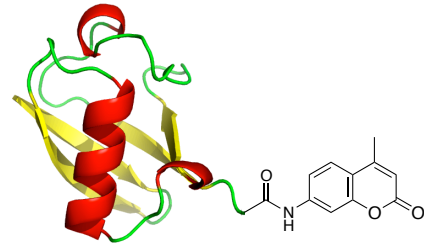
**Literature.** (1) El Oualid et al. *Angew Chem Int Ed* **2010**, 49, 10149. (2a) Dang et al. *Biochemistry* **1998**, 37, 1868. (2b) Mason et al. *Biochemistry* **2004**, 43, 6535. (3) Hassiepin et al. *Analyt Biochem* **2007**, 371, 201. (4a) Huang et al. *Methods in Molecular Biology* **2009**, 565, 127. (4b) Geurink et al. *ChemBiochem*, **2012**, 13, 293. (5a) White et al. *J Am Chem Soc* **1966**, 88, 2015. (5b) Reddy et al. *J Am Chem Soc* **2010**, 132, 13586. (5c) Orcutt et al. *Biochim Biophys Acta* **2012**, 1823, 2079.

# UbiQ

targeting the ubiquitin system

## Ub-AMC (human sequence, synthetic)

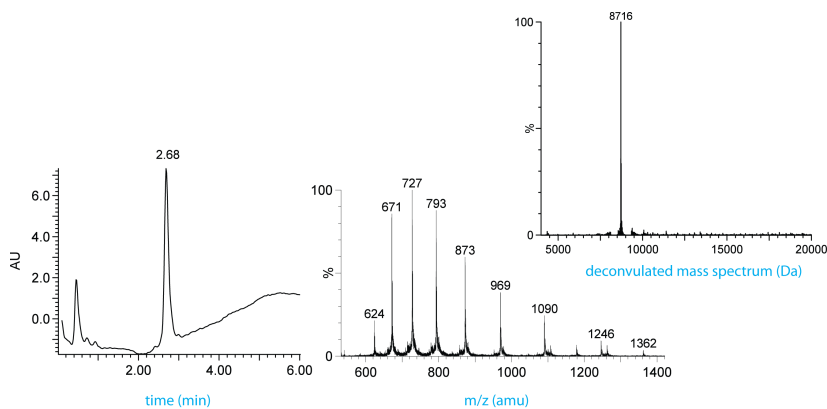
UbiQ code : UbiQ-001  
Batch # : B01012013-001  
Purity :  $\geq 95\%$  by RP-HPLC  
Amount : 25  $\mu\text{g}$ , lyophilized powder  
Mol. Weight : 8.72 kDa



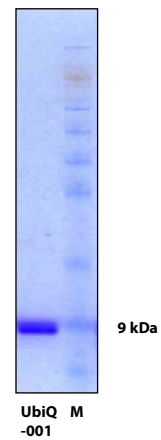
## sequence

**MQIFVKTLTGKTITLEVEPSDTIENVKAKIQDKEGIPPDQQRLIFAGKQLEDGRTLSDYNIQKESTLHLVLRLLGG-AMC**

**A**



**B**



**A: LC-MS analysis.** Mobile phase A = 1%  $\text{CH}_3\text{CN}$ , 0.1% formic acid in water (milliQ) and B = 1% water (milliQ) and 0.1% formic acid in  $\text{CH}_3\text{CN}$ . Phenomenex Kinetex C18, (2.1 $\times$ 50 mm, 2.6  $\mu\text{M}$ ); flow rate = 0.8 mL/min, runtime = 6 min, column T = 40 $^\circ\text{C}$ . Gradient: 5%  $\Rightarrow$  95% over 3.5 min.

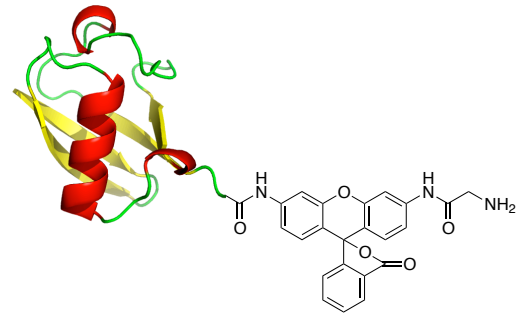
**B: SDS-PAGE analysis.** 12% SDS-PAGE gel. M= SeeBlue<sup>®</sup> Plus2 (Invitrogen). Coomassie Brilliant Blue staining.

# UbiQ

targeting the ubiquitin system

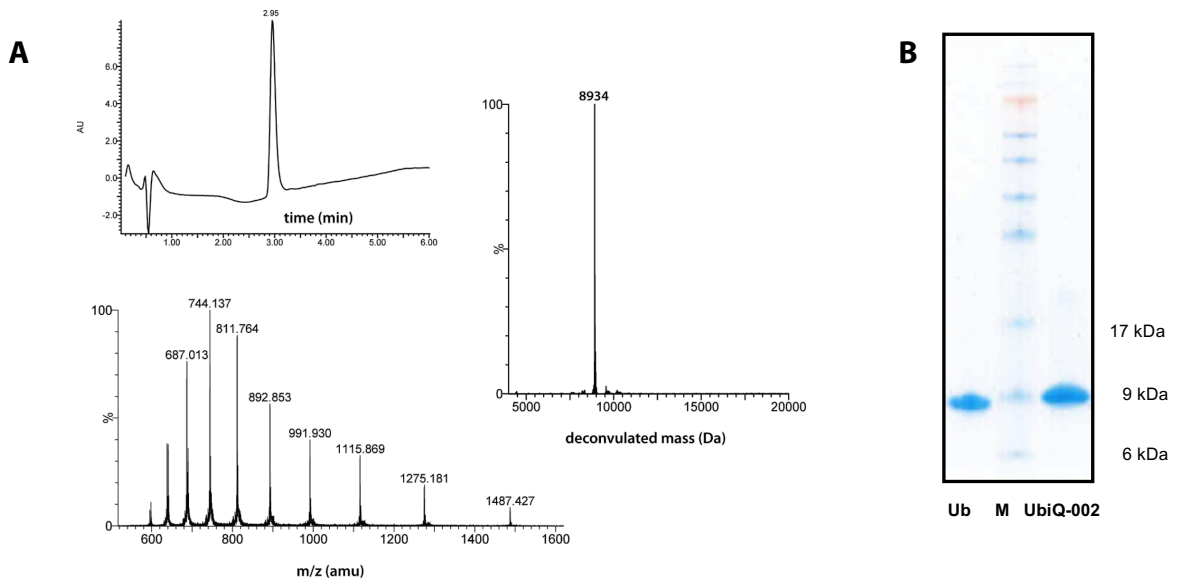
## Ub-Rh110Gly (human sequence, synthetic)

UbiQ code : UbiQ-002  
 Batch # : B01092013-001  
 Purity : ≥95% by RP-HPLC  
 Amount : 25 ug, lyophilized powder  
 Mol. Weight : 8.93 kDa



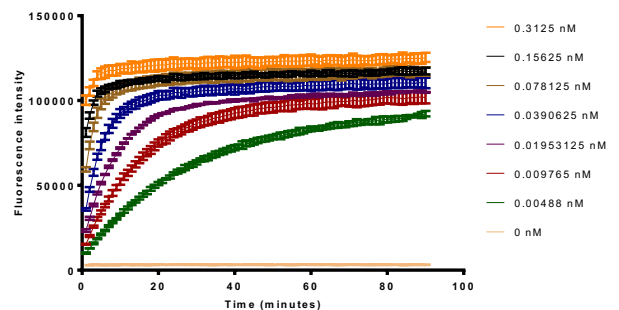
## sequence

**MQIFVKTLTGKTTITLEVEPSDTIENVKAKIQDKEGIPPDQORLIFAGKQLEDGRITLSDYNIQKESTLHLVLRRLGG-Rh110Gly**



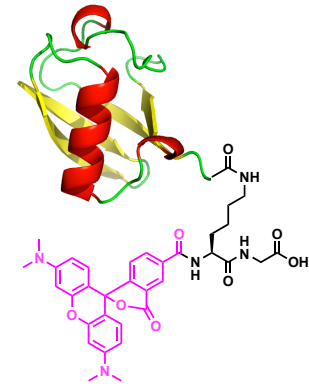
**A: LC-MS analysis.** Phenomenex Kinetex C18, (2.1×50 mm), 2.6 μM; flow rate = 0.5 mL/min, runtime = 6 min, column T = 40°C. Gradient: 5% ⇒ 95% B over 3.5 min. Mobile phase A = 1% CH<sub>3</sub>CN, 0.1% formic acid in water (milliQ) and B = 1% water (milliQ) and 0.1% formic acid in CH<sub>3</sub>CN. **B: SDS-PAGE analysis.** 12% SDS gel, MES buffer. Coomassie Brilliant Blue staining. M= SeeBlue® Plus2 (Invitrogen). **C: Activity assay.** 100 nM UbiQ-002 (B01092013-001) with varying concentrations of UCH-L3 (5 - 313 pM).

## C UbiQ-002 (B01092013-001) + UCH-L3



# UbiQ

targeting the ubiquitin system



## 5-TAMRA-Lys(Ub)-Gly-OH (human sequence, synthetic)

UbiQ code : UbiQ-012  
 Batch # : B24012013-001  
 Purity : ≥95% by RP-HPLC  
 Amount : 25 ug, lyophilized powder  
 Mol. Weight : 9.16 kDa

## sequence

5-TAMRA-Lys(MQIFVKTLTGKTITLEVEPSDTIENVKAKIQDKEGIPPDQQLIFAGKQLEDGRTLSDYNIQKESTLHLVLRRLRGG)-Gly

## Fluorescence polarization assay protocol

- FP assays were performed on a PerkinElmer Wallac EnVision 2010 Multilabel Reader with a 531 nm excitation filter and two 579 nm emission filters. Fluorescence intensities were measured in the S (parallel) and P (perpendicular) direction. FP values are given in mP (millipolarization) and calculated using formula (I)
- The confocal optics were adjusted with the average P and S values for TAMRA-Lys-Gly and the grating factor (G) was determined using a polarization value (L) for TAMRA of 50 mP using formula (II)
- The assays were performed in “non-binding surface flat bottom low flange” black 384-well plates (Corning) at room temperature in a buffer containing 20 mM Tris-HCl, pH 7.5, 5 mM DTT, 100 mM NaCl, 1 mg/mL 3-[(3-cholamidopropyl) dimethylammonio] propanesulfonic acid (CHAPS) and 0.5 mg/mL bovine gamma globulin (BGG). Each well had a volume of 20 μL. Buffer and enzyme were predispensed and the reaction was started by the addition of substrate. Kinetic data was collected in intervals of 2.5 or 3 min. From the obtained polarization values ( $P_t$ ) the amount of processed substrate ( $S_t$ ) was calculated with formula (III);  $P_t$  is the polarization measured (in mP);  $P_{max}$  is the polarization of 100% unprocessed substrate (determined for every reagent at all used substrate concentrations);  $P_{min}$  is the polarization of 100% processed substrate;  $S_0$  is the amount of substrate added to the reaction.
- From the obtained  $P_t$  values the values for initial velocities ( $v_i$ ) were calculated, which were used to determine the Michaelis-Menten constants ( $K_m$ ,  $V_{max}$  and  $k_{cat}$ ) by fitting the data according to formula (IV) (where  $k_{cat} = V_{max}/[E]$ ). All exp. data was processed using Ms Excel and Prism 4.03 (GraphPad Software, Inc).

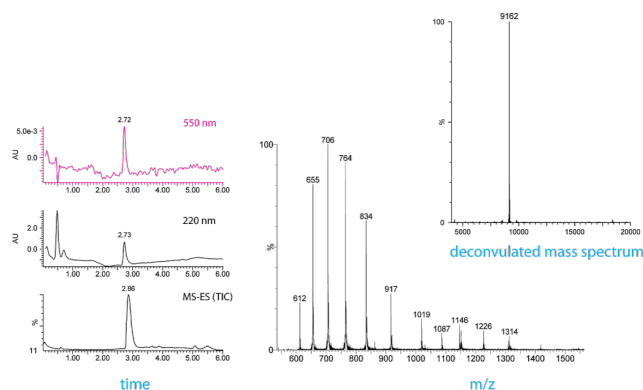
$$(I) \quad Polarization (mP) = \frac{S - (G \cdot P)}{S + (G \cdot P)} \cdot 1000$$

$$(II) \quad G = \frac{average\ S}{average\ P} \cdot \frac{1 - (\frac{L}{1000})}{1 + (\frac{L}{1000})}$$

$$(III) \quad S_t = S_0 \cdot \left[ S_0 \times \frac{P_t - P_{min}}{P_{max} - P_{min}} \right]$$

$$(IV) \quad v_i = \frac{V_{max} \times S_0}{K_m + S_0}$$

A



B



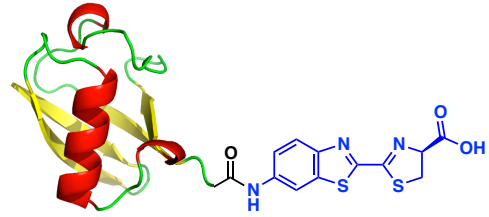
**A: LC-MS analysis.** Mobile phase A = 1% CH<sub>3</sub>CN, 0.1% formic acid in water (milliQ) and B = 1% water (milliQ) and 0.1% formic acid in CH<sub>3</sub>CN. Phenomenex Kinetex C18, (2.1×50 mm), 2.6 μm; flow rate = 0.5 mL/min, runtime = 6 min, column T = 40°C. Gradient: 5% ⇒ 95% over 3.5 min. **B: SDS-PAGE analysis.** 12% SDS-PAGE gel. CBB staining. M= SeeBlue® Plus2 (Invitrogen)

# UbiQ

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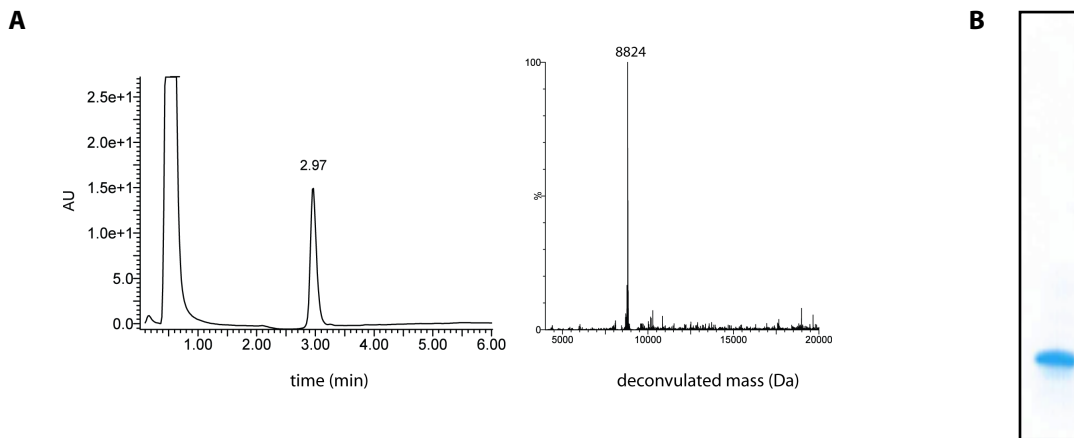
## Ubiquitin-aminoluciferin (human sequence, synthetic)

UbiQ code : UbiQ-036  
Batch # : B01092013-001  
Purity :  $\geq 95\%$  by RP-HPLC  
Amount : 25  $\mu\text{g}$ , lyophilized powder  
Mol. Weight : 8.82 kDa



## sequence

MQIFVKTLTGKTITLEVEPSDTIENVKAKIQDKEGIPPDQQLIFAGKQLEDGRTLSDYNIQKESTLHLVLRRLGG-(6-aminoluciferin)



**A: LC-MS analysis.** Mobile phase A = 1%  $\text{CH}_3\text{CN}$ , 0.1% formic acid in water (milliQ) and B = 1% water (milliQ) and 0.1% formic acid in  $\text{CH}_3\text{CN}$ . Phenomenex Kinetex C18, (2.1 $\times$ 50 mm), 2.6  $\mu\text{M}$ ); flow rate = 0.5 mL/min, runtime = 6 min, column T = 40°C. Gradient: 5%  $\Rightarrow$  95% over 3.5 min. **B: SDS-PAGE analysis.** 12% SDS-PAGE gel. Coomassie Brilliant Blue staining. M= SeeBlue® Plus2 (Invitrogen)